SYNTHESIS OF ANALOGUES OF 3-DEOXY-D-manno-OCTULOSONIC ACID (KDO) AS POTENTIAL INHIBITORS OF CMP-KDO SYNTHETASE

KRISTINA LUTHMAN, ALF CLAESSON*,

Department of Organic Pharmaceutical Chemistry, Uppsala Biomedical Center, University of Uppsala, P.O. Box 574, S-751 23 Uppsala (Sweden)

ANITA M. JANSSON, AND BRIAN G. PRING

Research and Development Laboratories, Antibacterial Chemotherapy, Astra Alab AB, S-151 85 Södertälje (Sweden)

(Received February 10th, 1987; accepted for publication, March 28th, 1987)

ABSTRACT

A series of derivatives of the 2-deoxy analogue of β -KDO (2,6-anhydro-3-deoxy-D-glycero-D-talo-octonic acid; ammonium salt, 2) has been synthesised as potential inhibitors of CMP-KDO synthetase, starting from methyl 2,6-anhydro-3-deoxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octonate and replacing the CO₂Me group attached to C-2 variously by CONH₂, CONHOH, CH₂OH, CH₂PO(OH)(O-NH₄+), COCH₂PO(OH)(O-H₃N \), CH₂CO₂-NH₄+, CON-HCH₂CO₂-NH₄+, CONHBn, CONHHexyl, CO₂Bn, and CO₂Hexyl. Of these derivatives, the hydroxamic acid (CONHOH) was the best inhibitor of CMP-KDO synthetase, but was less potent than 2.

INTRODUCTION

In a search for novel antibacterial agents with specificity for Gram-negative bacteria, a synthesis programme has been started aimed at potential inhibitors of enzymes involved in the metabolism of 3-deoxy-D-manno-octulosonic acid^{1,2} (KDO). KDO is a constituent of the lipopolysaccharide (LPS) of Gram-negative bacteria, which links the polysaccharide and lipid A, a di-(2-amino-2-deoxy-D-glucose) phosphate polymer^{3,4}. The biosynthesis of KDO and its incorporation into LPS is shown in Fig. 1. The choice of KDO metabolism as a target for antibacterial action is based on the facts that (a) KDO is found in Gram-negative bacteria⁵, possibly in the protozoan *Trypanosoma cruzi*⁶, and some higher plants⁷, (b) KDO is a component of the LPS of almost all Gram-negative bacteria examined⁵ and is

^{*}Present address: Research and Development Laboratorics, Antibacterial Chemotherapy, Astra Alab AB, S-151 85 Södertälje, Sweden.

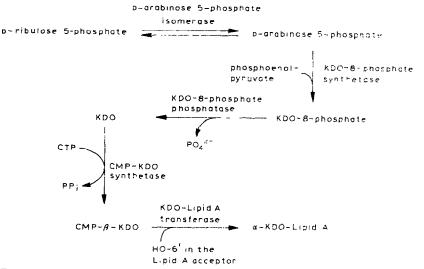


Fig. 1. Biosynthesis of KDO and its incorporation into LPS.

present also in the capsular polysaccharides of some strains⁸⁻¹⁵, and (c) mutants defective in KDO biosynthesis are not viable, and hence the inhibition of any relevant enzyme involved will result in inhibition of cell growth^{16,17}.

We have concentrated on the inhibition of CTP: 3-deoxy-p-manno-octulosonate cytidylyltransferase (CMP-KDO synthetase, EC 2.7.7.38)¹⁸⁻²⁰, which catalyses the formation of CMP-KDO from KDO and cytidine triphosphate (CTP). The activation of KDO before transfer to lipid A (Fig. 1) is believed to be the rate-limiting step in LPS biosynthesis¹⁰.

Substrate analogues of KDO have been synthesised as potential inhibitors of CMP-KDO synthetase. 5-Azido-5-deoxy-KDO and 5,8-diazido-5,8-dideoxy-KDO inhibited the enzyme², whereas 8-azido-8-deoxy-KDO and 4-O-methanesulfonyl-p-gluco-KDO were alternative substrates for the CMP-KDO synthetase-mediated reaction²¹. Compound 2, a deoxy analogue of β -KDO, is a competitive inhibitor of CMP-KDO synthetase, with a K_i of 3.9 μ M (ref. 22), whereas its epimer 3 is inactive. This observation indicates that β -KDO is the substrate for CMP-KDO synthetase, which accords with the finding, based on 13 C-n.m.r. studies, that the enzyme utilizes the β -pyranose form²³.

The enzyme-catalysed transfer of KDO to lipid A (Fig. 1) is thought to proceed with inversion of the anomeric configuration by analogy with CMP-N-acetylneuraminic acid (CMP-NANA)². Thus, KDO in native LPS ought to be α and this has been confirmed by n.m.r. experiments²⁴⁻²⁶.

We now report on a series of derivatives of 2 with variations of the $2-\alpha$ substituent, and their effects on CMP-KDO synthetase.

RESULTS AND DISCUSSION

The syntheses started with methyl 2,6-anhydro-3-deoxy-4,5:7,8-di-O-iso-propylidene-D-glycero-D-talo-octonate²⁷ (4), obtained from a derivative of 2-chloro-2-deoxy-α-KDO (ref. 28).

The dicarboxylate 9 was synthesised as follows. Reaction of the lithium enolate of 4 with carbon dioxide²⁹ afforded a 1:1 mixture of epimeric lithium carboxylates 5, which was converted into the diester 6 with iodomethane. The isopropylidene groups were hydrolysed from 6 with Amberlite IR-120 (H⁺) resin to afford 7. Saponification (0.2M NaOH, 25°) of 7 gave the half-ester 8, and further treatment (2M NaOH, 60°) afforded 9, without detectable decarboxylation, and which was isolated as the diammonium salt. The amide 10 was easily obtained from 4 by treatment with saturated methanolic ammonia and deprotection then gave 11. The hydroxamic acid 12 was prepared by reaction of 4 with hydroxylamine hydrochloride in methanol in the presence of triethylamine.

Reduction of 4 with LiAlH₄ gave the alcohol 14 which, with triphenylphosphine and N-bromosuccinimide, gave the bromide 16. An Arbuzov reaction in triethyl phosphite then afforded 17. Treatment of 17 with bromotrimethylsilane in chloroform³⁰ effected transesterification to give the bis(trimethylsilyl)phosphonate

34 R = CO2Hexy!

that was cleaved by water to yield the phosphonic acid, which catalysed acetal hydrolysis to give 18, isolated as the ammonium salt.

The β -ketophosphonate 19 was obtained from 4 and lithium dimethyl methylphosphonate³¹ and deprotected to give 20 isolated as the cyclohexylammonium salt.

The homocarboxylic acid analogue 24 was synthesised as follows. The tosylate 21 of 14 was treated with KCN to give the nitrile 22. Acid-catalysed methanolysis of 22 with chlorotrimethylsilane as a water scavenger afforded the methyl ester 23, which was saponified to yield 24, isolated as the ammonium salt. The glycine derivative 27 was obtained via 26, which was prepared from a mixed anhydride of the carboxylate 25 and glycine methyl ester hydrochloride in the presence of triethylamine. Likewise, the benzyl- (29) and hexyl-amide (31) were obtained via 28 and 30, respectively, which were synthesised from 25. The esters 33 and 35 were prepared by deprotection of 32 and 34, respectively, which were synthesised by transesterification of 4 with catalytic amounts of magnesium methoxide in benzyl alcohol or 1-hexanol, respectively. Magnesium methoxide was used instead of sodium methoxide in order to avoid epimerisation during the transesterification step.

N.m.r. data. — The structures of the foregoing compounds were assigned on the basis of ¹H-, ¹³C-, and ²¹P-n.m.r. data. The ¹³C-n.m.r. spectra were proton noise-decoupled and the signals of **10** were assigned by using a graphical method involving proton off-resonance spin-decoupled spectra^{32,33}. The ¹³C-n.m.r. spectra

of 12, 26, 28, and 30 were interpreted by comparison with data for 10, since variations in the 2-substituent had little effect on the ¹³C resonances.

The configuration at C-2 in **8** was determined by analogy with a method used for determination of the anomeric configurations in CMP-NANA (ref. 34) and KDO-glycosides^{25,35}. The J values for the carbonyl carbons and H-3,3 provided evidence for the configuration at C-2. The resonances of the carbonyl carbons were assigned on the basis of the respective multiplets in the proton-coupled ¹³C-n.m.r. spectra. The carboxylate signal appeared as a doublet (${}^{3}J_{\text{C,H-3}a}$ 3 Hz) and the ester carbonyl signal as an unresolved multiplet. Selective ¹³C-[¹H] decoupling centered at the chemical shift of the methyl ester protons at 3.74 p.p.m. collapsed the multiplet at 174.82 p.p.m. to a doublet of doublets with ${}^{3}J_{\text{C,H-3}a}$ 9 Hz, and ${}^{3}J_{\text{C,H-3}e}$ 3 Hz, thereby supporting the structure assigned to **8**.

The 400-MHz ¹H-n.m.r. spectra indicated the amide **10** to exist in solution preponderantly in the distorted boat conformation **36**. The 5C_2 chair conformation is ruled out because the coupling constants $J_{2,3a}$ 11.6 and $J_{2,3e}$ 5.8 Hz are too large to account for an axial-equatorial and equatorial-equatorial relationship, respectively. In addition, the value $J_{3a,4}$ 2.7 Hz does not correspond to a diaxial coupling. However, the large value (11.6 Hz) of $J_{2,3a}$ indicated a *trans*-diaxial relationship of H-2,3a. Thus, H-2 must occupy an axial or pseudo-axial position. The alternative 2C_5 chair conformation is unlikely due to a considerable 4,6-diaxial interaction.

The J values for 10 are compared in Table I with data determined for derivatives of 1,2:3,4-di-O-isopropylidene- α -D-galactose^{36,37}. Variations in the structure of the side chain do not affect the J values for the ring protons. When comparing experimental and calculated J values, Cone and Hough³⁶ established that the galactose derivatives must adopt a conformation intermediate between skew and boat forms. The J values for 10 and the other O-isopropylidene derivatives accord with these results (Table I). Recently, Krajewski et al.³⁸ reported that the pyranose ring in 6-C-(2-furyl)-1,2:3,4-di-O-isopropylidene- α -D-glycero-D-galacto-hexopyranose adopted a hybrid twist-boat conformation (${}^{\circ}T_2 + B_{2,5}$), thus further supporting the conformation proposed for 10. However, the pyranose ring in crystalline 3,4,5-tri-O-acetyl-1,2-O-(R)-(1-cyanoethylidene)- α -D-galactopyranose adopts a flattened ${}^{4}C_1$ conformation³⁹. The values $J_{2,3}$ 7.4 and $J_{3,4}$ 3.0 Hz for compound C (Table I) do not accord with those for the other derivatives in Table I. Thus, the conformational restraints imposed on the pyranose ring by the 3,4-O-iso-

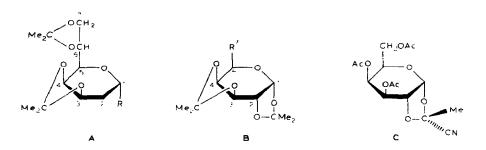


TABLE I
COUPLING CONSTANTS⁴

Compound	J _{1.,2a}	J _{1.2e}	J _{2a, t}	$J_{\lambda_{e,i}}$	J.,	J ₄₅	J _{5 6}
Series A							
$4 R = CO_2 Me^{30}$	11.8	5.9	2.7	3.2	8.0	1.5	7.8
$10 R = CONH_2$	11.6	5.8	2.7	2.8	8.0	1.5	4.3
12 R = CONHOH	11.8	6.0	2.5	2.8		1.5	4.0
		2.9	2.9	8.3	1.5		7.2
$19 R = COCH_2PO(OMe)_2$	11.4	5.1	2.7	2.7	8.1	1.7	6.8
$21 R = CH_2OTs$			3.0	3.0	8.0	1.5	8.0
$22 R = CH_2CN$			3.0	3.0	8.0	1.5	8.0
$25 R = CO_2 - TEA$	11.0	6.0	3.0	3.2	8.0	1.5	8.0
$26 R = CONHCH_2CO_2Me$	11.8	5.5	2.5	2.8		1.5	4.8
28 R = CONHBn	11.6	6.0	2.6	3.0		1.5	3.5
$30 R = CONHC_6H_{13}$	11.7	6.0	2.5	3.0	8.0	1.5	4.0
$32 R = CO_2Bn$	11.7	5.9	2.4	3.4	7.8	1.5	8.3
$34 R = CO_2C_6H_{13}$	11.7	5.9	2.4	3.2	7.8	1.5	8.0
Series B							
R' = H		5.0		2.4	7.8	1.4; 2.0	
R' = COSBn		5.1		2.4	8.0	1.4	
$R' = CH_2I$		5.0		2.4	7.8	1.6	5.9; 7.7
$R' = CH_2OH$		5.0		2.4	8.0	1.4	
$R' = CH_2OTs$		5.0		2.4	7.8		
Compound C		4.6		7.4	3.0	2.4	

"For the 4.5:7,8-di-O-isopropylidene derivatives (A), 1,2:3,4-di-O-isopropylidenegalactose derivatives $^{16.37}$ (B), and 3,4,6-tri-O-acetyl-1,2-O-(R)-(1-cyanocthylidene)- α -D-galactopyranose 19 (C) (for convenience, all derivatives are numbered in the same way).

propylidene ring dominate those of the same ring in the 1,2-positions.

If, in solution, a rapid interconversion between the skew and boat forms of 10 occurs, the observed chemical shifts and coupling constants will be weighted averages. However, determination of the ${}^{1}\text{H-n.m.r.}$ spectra at -40° and -60° did not change the chemical shifts and coupling constants, suggesting that 10 exists in only one conformation. It is possible that 10 exists in two conformations with a low barrier of interconversion. In the deduced conformation of 10, the bulky side-chain at C-6 occupies the more stable equatorial position. Studies of molecular models reveal that a diaxial relationship of H-6,7 corresponds to the most stable conforma-

TABLE II
RELATIVE INHIBITORY ACTIVITY IN THE CMP-KDO SYNTHETASE ASSAY

Compounda	Relative activity ^{b-d}	
2	++++	
8	Ī	
9	I	
11	++	
13	+++	
15	I	
18	Ī	
20	+	
24	Ī	
25	I	
27	+	

Compounds 29, 31, 32, and 35 were inactive. ^bEquimolar concentrations of KDO and inhibitor. ^cKey: I, inactive; +, <50% inhibition; ++, 50–80%; ++++, 80–90%; +++++, 90–99%; ++++++, 100%. ^dAll compounds tested were analysed by h.p.l.c.⁴¹ to ensure the absence of contamination by 2.

tion, due to minimal steric interactions with the ring. However, the small $J_{6.7}$ value of 4 Hz for 10 suggests a gauche relationship. Approximately the same $J_{6.7}$ value was found for 12, 26, 28, and 30 (Table I). This side-chain conformation might be due to the formation of a hydrogen bond between NH and O-7, which is not possible in 21, 22, 25, 32, and 34 where $J_{6.7}$ is \sim 8 Hz, a value consistent with a diaxial coupling. The presumed hydrogen bond between NH and O-7 could not be disrupted even in a methanolic solution of 10 at 55°.

Biological activity. — The compounds described above were tested as inhibitors of CMP-KDO synthetase from Escherichia coli D21 or a mutant of Salmonella typhimurium (SL 1102). The CMP-KDO synthetase screening-assay was based essentially on that described by Ghalambor and Heath¹⁸. The KDO liberated from the nucleotide was determined by a modified thiobarbituric acid assay⁴⁰. All of the compounds were less potent than 2 (Table II). The most potent was 13, followed by 11. The β -ketophosphonate 20 and the glycine analogue 27 were weak inhibitors, and the dicarboxylate 9, the phosphonate 18, and the homocarboxylate 24 were inactive. The inhibitory activity of the hydroxamic acid 13, which is a weak acid, is probably due to the location of a negative charge at the same relative spatial position as that in 2. However, a negative charge is not absolutely essential since the amide 11 was also a rather good inhibitor. Thus, the polarity of a correctly positioned amide group appears to be sufficient for activity. The importance of an appropriate spatial location of the polar group is shown by moving the anionic centre further from the ring, as in compounds 18, 20, 24, and 27 which were much less potent as inhibitors. Compound 9 was inactive even though the negative charge is at the same position as in 2. This effect might be due to the equatorial carboxylate group preventing simultaneous binding of 9 and CTP to the active site of the enzyme²².

The alcohol 15 and the carboxylates 8 and 25, which are intermediates in the synthetic pathways, were also inactive.

In order to investigate whether penetration of the bacterial cell wall might be improved, some hydrophobic derivatives of 2 were synthesised, namely, the amides 29 and 31 and the esters 33 and 35, with the idea that intracellular hydrolysis might liberate the potent inhibitor 2. However, these compounds, in addition to 11, 13, 25, and 27, did not inhibit the growth⁴² of Salmonella typhimurium SL 1102, Salmonella typhimurium LT2 MI, and Escherichia coli ATCC 11303 at 37° and Salmonella typhimurium AG701i50 at 25°.

EXPERIMENTAL

General methods. — Melting points were determined in open capillary tubes and are uncorrected. Optical rotations were measured at 20° with a Perkin–Elmer 241 or Optical Activity AA100 polarimeter. I.r. spectra were recorded with a Perkin–Elmer 298 spectrometer. N.m.r. spectra were recorded with a JEOL FX90Q, JEOL FX200, or JEOL GX-400 instrument on solutions in CDCl₃ and CD₃OD (internal Me₄Si), D₂O (internal BuOH, $\delta_{\rm H}$ 1.23, $\delta_{\rm C}$ 32.2), or as indicated otherwise. H₃PO₄ (δ 0) was used as external standard for ³¹P-n.m.r. spectra. High-resolution f.a.b.-mass spectra were recorded with a JEOL DX 303 instrument. T.l.c. was performed on Merck Silica Gel 60 F₂₅₄ with detection by u.v. light and by charring with sulphuric acid. Column chromatography was performed on Merck Silica Gel 60 (0.040–0.063 mm), using the flash technique⁴³. All solvents used were anhydrous and kept over 3Å and 4Å molecular sieves. N,N-Dimethylformamide was distilled from P₂O₅ and then kept over 4Å molecular sieves. Butyl-lithium in hexane was titrated prior to use⁴⁴. Solutions were concentrated *in vacuo* at <30°.

Hydrolysis of di-O-isopropylidene derivatives. — A solution of the di-O-isopropylidene derivative in methanolic 20% trifluoroacetic acid was stirred at room temperature until hydrolysis was complete (t.l.c.) and then co-concentrated several times with toluene, and the residue was recrystallised or purified as indicated.

Methyl 2,6-anhydro-3-deoxy-4,5:7,8-di-O-isopropylidene-2-methoxycarbonyl-D-glycero-D-talo-octonate (6). — 1.6M Butyl-lithium in hexane (3.4 mL, 5.4 mmol) was added to a solution of di-isopropylamine (560 mg, 5.5 mmol) in dry tetrahydrofuran (20 mL) at -20° under nitrogen. After 20 min, the mixture was cooled to -75° and a solution of 4 (1.70 g, 5.4 mmol) in dry tetrahydrofuran (5 mL) was added dropwise. Carbon dioxide was bubbled through the solution for 15 min, the temperature was slowly raised to 0°, and, after the addition of water (17 mL), the solution was neutralised with Dowex (H⁺) resin, washed three times with chloroform, and concentrated to give 5 (1.80 g, 95%) as a white powder, $R_{\rm F}$ 0.70 (chloroform–methanol–water, 10:10:1); $\nu_{\rm max}$ (CHCl₃), 1730 (ester), 1640 and 1400 cm⁻¹ (carboxylate). ¹H-N.m.r. data (CDCl₃): δ 3.70 and 3.72 (OMe). The n.m.r. data indicated this product to be a 1:1 mixture (5) of lithium 2.6-anhydro-3-deoxy-4,5:7,8-di-*O*-isopropylidene-2-methoxycarbonyl-D-glycero-D-talo- and -D-galacto-octonate, which was used as such in the next step.

Cesium carbonate (1.90 g, 5.8 mmol) and iodomethane (0.82 mL, 13 mmol) were added to a solution of 5 (1.80 g, 4.9 mmol) in dry tetrahydrofuran (25 mL). After 2 h at room temperature, ethyl acetate (25 mL) was added, and the mixture was filtered and concentrated. Column chromatography (ethyl acetate–toluene, 1:2) of the residue gave 6 (1.43 g, 78%), m.p. $103-104^{\circ}$ (from ethyl acetate–hexane), $[\alpha]_D$ -6° (c 0.3, dichloromethane), R_F 0.38 (ethyl acetate–toluene, 1:2); $\nu_{\rm max}$ (CHCl₃) 1735 cm⁻¹ (ester). N.m.r. data (CDCl₃): 1 H, δ 1.34, 1.36, 1.42 (3 s, 12 H, 2 Me₂C), 2.25 (dd, 1 H, $J_{3a,3e}$ -15.6, $J_{3a,4}$ 2.2 Hz, H-3a), 2.95 (dd, 1 H, $J_{3e,4}$ 3.9 Hz, H-3e), 3.49 (dd, 1 H, $J_{6,7}$ 8.3, $J_{6,5}$ 1.5 Hz, H-6), 3.78 (s, 6 H, 2 OMe), 4.05–4.65 (m, 5 H, H-4,5,7,8,8'); 13 C, δ 24.86, 25.08, 25.34, 27.07 (2 CMe₂), 28.92 (C-3), 52.92, 53.17 (2 OMe), 67.23 (C-8), 69.47, 72.02, 73.48, 74.07 (C-4,5,6,7), 78.57 (C-2), 109.41, 109.70 (2 CMe₂), 169.20, 169.52.

Anal. Calc. for C₁₇H₂₆O₉: C, 54.5; H, 7.0. Found: C, 54.55, H, 7.1.

Methyl 2,6-anhydro-3-deoxy-2-methoxycarbonyl-D-glycero-D-talo-octonate (7). — A mixture of 6 (340 mg, 0.91 mmol), Amberlite IR-120 (H⁺) resin (2 mL, prewashed with anhydrous methanol), and anhydrous methanol (12 mL) was heated under reflux for 2 h, then filtered, and concentrated. Column chromatography (ethyl acetate-methanol-toluene, 14:2:1) of the residue gave 7 (210 mg, 77%), m.p. 145–148° (from methanol-ether), [α]_D +77° (c 0.2, methanol), R_F 0.30 (ethyl acetate-methanol-toluene, 7:2:1); ν_{max} (CHCl₃) 1740 cm⁻¹ (ester). N.m.r. data (CD₃OD): ¹H, δ 2.08 (dd, 1 H, $J_{3a,3e}$ –12.7, $J_{3a,4}$ 12.7 Hz, H-3a), 2.38 (dd, 1 H, $J_{3e,4}$ 4.6 Hz, H-3e), 3.35 (d, 1 H, $J_{6,7}$ 8.1 Hz, H-6), 3.59–3.93 (m, 11 H, H-4,5,7,8,8' and OMe at δ 3.76 and 3.78); ¹³C, δ 32.84 (C-3), 53.64, 53.73 (2 OMe), 65.17 (C-8), 67.28, 67.96 (C-4,5), 70.81, 77.42 (C-6,7), 83.17 (C-2), 169.62, 170.20 (2 C=O).

Anal. Calc. for $C_{11}H_{18}O_9$: C, 44.9; H, 6.2. Found: C, 44.5; H, 6.3.

Ammonium 2,6-anhydro-3-deoxy-2-methoxycarbonyl-D-glycero-D-galacto-octonate (8). — A solution of 7 (240 mg, 0.81 mmol) in 0.2M NaOH (10 mL) was stored at room temperature for 45 min, then passed through Dowex 50W-X8 (NH⁺₄) resin, and concentrated to give 8 (240 mg, 100%), as a semi-crystalline product after prolonged storage in the refrigerator, m.p. 150° (dec.), $[\alpha]_D$ +107° (c 1.5, water), R_F 0.35 (chloroform-methanol-water, 10:10:1); $\nu_{\text{max}}^{\text{KBr}}$ 1730 (ester), 1600 cm⁻¹ (carboxylate). N.m.r. data (D₂O): ¹H, δ 1.84 (dd, 1 H, $J_{3a,3e}$ –13.1, $J_{3a,4}$ 13.1 Hz, H-3a), 2.49 (dd, 1 H, $J_{3e,4}$ 4.8 Hz, H-3e), 3.21 (d, 1 H, $J_{6,7}$ 8.3 Hz, H-6), 3.6–3.94 (m, 8 H, H-4,5,7,8,8′, OMe at δ 3.74); ¹³C, δ 34.10 (C-3), 55.70 (OMe), 65.94 (C-8), 68.54, 69.42, 71.87, 77.83 (C-4,5,6,7), 86.37 (C-2), 174.82 (ester C=O), 176.33 (carboxylate C=O).

Anal. Calc. for $C_{10}H_{19}NO_9 \cdot H_2O$: C, 38.1; H, 6.7; N, 4.4. Found: C, 37.8; H, 6.9; N, 4.45.

Diammonium 2,6-anhydro-3-deoxy-2-carboxy-D-glycero-D-talo-octonate (9). — Hydrolysis of 8 (150 mg, 0.5 mmol) in 2M NaOH (2 mL) at 60° for 20 min gave, after passage through Dowex 50W-X8 (NH₄) resin and concentration, 9 (150 mg, 100%), which crystallised on storage, m.p. 140° (dec.), $[\alpha]_D + 100^{\circ}$ (c 2, water);

 $\nu_{\text{max}}^{\text{KBr}}$ 1590–1600 cm⁻¹ (carboxylates). N.m.r. data (D₂O): ¹H, δ 1.70 (dd, 1 H, $J_{3a,3e}$ –12.7, $J_{3a,4}$ 12.7 Hz, H-3a), 2.50 (dd, 1 H, $J_{3e,4}$ 4.4 Hz, H-3e), 3.24 (d, 1 H, $J_{6,7}$ 7.8 Hz, H-6), 3.72–3.95 (m, 5 H, H-4,5,7,8.8'); ¹³C, δ 35.36 (C-3), 66.55 (C-8), 68.78, 70.22, 71.70, 76.93 (C-4,5.6,7), 87.90 (C-2), 178.64, 179.22 (C=O).

Anal. Calc. for $C_0H_{20}N_2O_9 \cdot H_2O \cdot C$, 34.0; H, 7.0; N, 8.8. Found: C, 34.2; H. 6.7; N, 8.7.

2,6-Anhydro-3-deoxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octonamide (10). — A solution of 4 (490 mg, 1.48 mmol) in saturated methanolic ammonia (10 mL) was stirred at room temperature for 2 days, and then concentrated. Recrystallisation of the residue from chloroform-pentane gave 10 (317 mg, 71%), m.p. 129–131°, $[\alpha]_D$ –38° (c 1.8, chloroform), R_f 0.76 (ether-acctone, 2:1). N.m.r. data (CDCl₃): 1H (400 MHz), δ 1.35, 1.39, 1.43, 1.47 (4 s, 12 H, 2 Me₂C), 1.80 (ddd, 1 H, $J_{3a,3e}$ –15.5, $J_{3a,2}$ 11.6, $J_{3a,4}$ 2.7 Hz, H-3a), 2.50 (ddd, 1 H, $J_{3c,2}$ 5.8, $J_{3c,4}$ 2.8 Hz, H-3e), 3.72 (dd, 1 H, $J_{6,7}$ 4.3, $J_{6,5}$ 1.5 Hz, H-6), 4.06–4.10 (m, 2 H, H-8.8'), 4.23–4.27 (m, 2 H, H-5,7), 4.48 (dd, 1 H, H-2), 4.61 (ddd, 1 H, $J_{4,5}$ 8.0 Hz, H-4), 5.8, 6.9 (NH₂); 13 C, δ 24.53, 25.47, 25.97, 26.47 (2 CMe₂), 26.97 (C-3), 65.36 (C-8), 69.24 (C-2), 70.24 (C-4), 71.39 (C-6), 72.28 (C-7), 75.38 (C-5), 108.38, 109.22 (2 CMe₂), 176.42 (C-1).

Anal. Calc. for C₁₄H₂₃NO₆: C, 55.8; H, 7.7; N, 4.65. Found: C, 56.0; H, 7.9; N, 4.4.

2,6-Anhydro-3-deoxy-D-glycero-D-talo-octonamide (11). — Compound 10 (200 mg, 0.66 mmol) was deprotected according to the standard procedure, to give 11 (145 mg, 98%). Recrystallisation from methanol-2-propanol gave material with m.p. 191-193°, $[\alpha]_D$ +71° (c 0.7, methanol-water 2:1). R_1 0.56 (chloroform-methanol-water, 10:10:1). N.m.r. data (D₂O): 1 H, δ 2.00 (ddd, 1 H, $J_{3a,3c}$ =12, $J_{3a,4}$ 12, $J_{3a,2}$ 6.5 Hz, H-3a), 2.25 (ddd, 1 H, $J_{3c,4}$ 5.5, $J_{3c,2}$ 1.5 Hz, H-3e), 3.52 (dd, 1 H, $J_{6,7}$ 8, $J_{6,5}$ 1 Hz, H-6), 3.66-4.0 (m, 5 H, H-4.5,7,8.8′), 4.58 (dd. 1 H, H-2); 13 C, δ 27.38 (C-3), 64.22 (C-8), 67.31, 67.39 (C-4.5), 70.15 (C-7), 74.44 (C-6), 75.44 (C-2), 177.18 (C-1).

Anal. Calc. for $C_8H_{15}NO_6$: C, 43.4; H, 6.8; N, 6.3. Found: C, 43.1; H, 6.8; N, 6.3.

2,6-Anhydro-3-deoxy-N-hydroxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octonamide (12). — Hydroxylamine hydrochloride (440 mg, 6.33 mmol) was added to a solution of 4 (400 mg, 1.26 mmol) and triethylamine (1.3 mL) in methanol (10 mL). The mixture was heated under reflux under nitrogen for 2 h, then stirred at room temperature overnight, and concentrated. Column chromatography (ether-acetone, 2:1) of the residue gave 12 (240 mg, 60%), isolated as a colourless syrup, [α]_D =31° (c 1, chloroform), R_F 0.75 (ether-acetone, 2:1). N.m.r. data (CDCl₃): 1 H, δ 1.26, 1.34, 1.39, 1.45 (4 s, 12 H, 2 Me₂C), 1.78 (ddd, 1 H, $J_{3a,3e}$ =15, $J_{3a,2}$ 11.8, $J_{3a,4}$ 2.5 Hz, H-3a), 2.48 (ddd, 1 H, $J_{3e,2}$ 6, $J_{3e,4}$ 2.8 Hz, H-3e), 2.63 (s, 1 H, OH), 3.71 (dd, 1 H, $J_{6,7}$ 4, $J_{6,5}$ 1.5 Hz, H-6), 3.95–4.35 (m, 4 H, H-5.7.8.8′), 4.50–4.75 (m, 2 H, H-2.4), 9.40 (s, 1 H, NH); 13 C, δ 24.52, 25.45, 25.94, 26.37 (2 CMe₂), 26.93 (C-3), 65.10 (C-8), 68.93 (C-2), 70.17 (C-4), 71.40 (C-6), 72.27 (C-7), 75.29 (C-5), 108.46, 109.39 (2 CMe₂), 169.92 (C-1).

Anal. Calc. for $C_{14}H_{23}NO_7 \cdot 0.5 H_2O$: C, 51.5; H, 7.4; N, 4.3. Found: C, 51.9; H, 7.2; N, 3.8.

2,6-Anhydro-3-deoxy-N-hydroxy-D-glycero-D-talo-octonamide (13). — Deprotection of 12 (123 mg; 0.4 mmol) by the standard procedure and crystallisation of the product from methanol-ether gave 13 (75 mg, 83%), m.p. 148–151°, $[\alpha]_D$ +76° (c 1, methanol), R_F 0.57 (chloroform-methanol-water, 10:10:1). N.m.r. data (D₂O): ¹H, δ 2.00 (ddd, 1 H, $J_{3a,3e}$ –12, $J_{3a,4}$ 12, $J_{3a,2}$ 6 Hz, H-3a), 2.25 (ddd, 1 H, $J_{3e,4}$ 6, $J_{3e,2}$ 2 Hz, H-3e), 3.46 (d, 1 H, $J_{6,7}$ 8 Hz, H-6), 3.57–4.05 (m, 5 H, H-4,5,7,8,8'), 4.62 (dd, 1 H, H-2); ¹³C, δ 28.80 (C-3), 65.67 (C-8), 68.52 (overlapping signals C-4,5), 71.29 (C-7), 74.94 (C-6), 76.86 (C-2), 172.10 (C-1).

Anal. Calc. for $C_8H_{15}NO_7$: C, 40.5; H, 6.4; N, 5.9. Found: C, 40.6; H, 6.4; N, 5.5.

2,6-Anhydro-3-deoxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octitol (14). — A solution of 4 (2.2 g, 6.9 mmol) in dry ether was added dropwise during 5 min to a suspension of LiAlH₄ (270 mg, 7.1 mmol) in dry ether (100 mL) under nitrogen. The mixture was heated under reflux for 35 min, water (1.6 mL) and 2M NaOH (0.7 mL) were then added, and the ether layer was dried (K_2CO_3), filtered, and concentrated to give 14 (2.0 g, 100%). Recrystallisation from ether-light petroleum gave material with m.p. 59-63°, [α]_D +0.7° (c 0.7, dichloromethane), R_F 0.40 (ethyl acetate). N.m.r. data (CDCl₃): 1 H, δ 1.38, 1.42, 1.50 (3 s, 12 H, 2 CMe₂), 1.7-1.9 (m, 2 H, H-3 α ,3e), 3.4-3.7 (m, 3 H, H-6, CH₂), 4.0-4.3 (m, 4 H, H-5,7,8,8'), 4.32 (dd, 1 H, $J_{5,4}$ 8.3, $J_{5,6}$ 1.5 Hz, H-5), 4.59 (ddd, 1 H, $J_{4,3a}$ 2.9, $J_{4,3e}$ 2.9 Hz, H-4): 13 C, δ 24.45, 25.27, 26.13, 26.88 (2 CMe₂), 26.88 (C-3), 65.56, 66.87 (C-1,8), 69.16 (C-2), 70.20, 71.52, 72.37, 74.58 (C-4,5,6,7), 108.81, 109.10 (2 CMe₂).

Anal. Calc. for C₁₄H₂₄O₆: C, 58.3; H, 8.4. Found: C, 58.2; H, 8.2.

2,6-Anhydro-3-deoxy-D-glycero-D-talo-octitol (15). — Compound 14 (300 mg, 1 mmol) was heated under reflux with Amberlite IR-120 (H⁺) resin (3 mL, prewashed with methanol and dried) in anhydrous methanol (10 mL) for 6 h. The mixture was then filtered and concentrated. Column chromatography (ethyl acetate-methanol-toluene, 65:25:10) of the residue gave 15 (100 mg, 50%), m.p. 93–94°, $[\alpha]_D$ +51° (c 1, methanol), R_F 0.29 (ethyl acetate-methanol-toluene, 13:5:2). N.m.r. data (CD₃OD): ¹H, δ 1.60 (dd, 1 H, $J_{3e,3a}$ –13, $J_{3e,2}$ 5 Hz, H-3e), 2.0 (ddd, 1 H, $J_{3a,2}$ 7, $J_{3a,4}$ 12 Hz, H-3a), 3.34–4.1 (m, 9 H, H-2,4,5,6,7,8,8', CH₂); ¹³C, δ 29.41 (C-3), 61.88 (C-1), 65.17 (C-8), 67.57, 68.77, 71.25, 72.78 (C-4,5,6,7), 74.43 (C-2).

Anal. Calc. for C₈H₁₆O₆: C, 46.15; H, 7.75. Found: C, 45.75; H, 7.5.

Monoammonium 2,6-anhydro-1,3-dideoxy-D-glycero-D-talo-octitol-1-ylphosphonate (18). — A mixture of N-bromosuccinimide (258 mg, 1.45 mmol), triphenylphosphine (380 mg, 1.45 mmol), 14 (380 mg, 1.32 mmol), and chloroform (10 mL) was heated under reflux for 1 h. More triphenylphosphine (380 mg) and N-bromosuccinimide (258 mg) were then added, and the solution was heated overnight, cooled, diluted with light petroleum, filtered, and concentrated. Column

chromatography (ether–light petroleum, 2:3) of the residue gave 2.6-anhydro-1-bromo-1,3-dideoxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octitol (**16**: 230 mg, 50%), $R_{\rm F}$ 0.67 (ethyl acetate). ¹³C-N.m.r. data (CDCl₃): δ 24.47, 25.27, 26.15, 26.98 (2 CMe₂), 29.06 (C-1), 37.29 (C-3), 67.21, 67.36, 70.15, 72.15, 72.61, 74.14 (C-2,4.5,6,7,8), 109.27, 109.98 (2 CMe₂).

Compound **16** (230 mg, 0.65 mmol) was heated under reflux with freshly distilled triethyl phosphite (10 mL) under nitrogen for 40 h and the solution was then concentrated at 80°/0.5 mmHg. Column chromatography [ethyl acetate-hexane, 1:4 (1 L); ethyl acetate-hexane, 1:1 (750 mL); and ethyl acetate (750 mL)] gave diethyl 2,6-anhydro-1,3-dideoxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octitol-1-ylphosphonate (**17**; 160 mg, 60%), $R_{\rm F}$ 0.14 (ethyl acetate). N.m.r. data (CDCl₃): 1 H, δ 1.2–1.6 (m, 18 H, including 2 CMe₂ at 1.32, 1.35, 1.41 and 1.49, and 2 ethyl Me), 1.8–2.2 (m, 4 H, H-1a,1b,3a,3e), 3.42 (d, 1 H, $J_{\rm 6,7}$ 8 Hz, H-6), 4.0–4.6 (m, 10 H, H-2,4,5.7,8,8', OC $H_{\rm 2}$ CH₃); 13 C, δ 16.45 (CH₃, $^{3}J_{\rm C,P}$ 6 Hz), 24.64, 25.27, 26.27, 27.07 (2 CMe₂), 31.64 (C-3, $^{3}J_{\rm C,P}$ 7 Hz), 33.24 (C-1, $^{1}J_{\rm C,P}$ 129 Hz), 61.55, 61.69 (2 CH₂, $^{2}J_{\rm C,P}$ 6 Hz), 64.84, 67.13, 70.29, 71.32, 72.29, 74.28 (C-2,4,5,6,7,8), 108.90, 109.24 (2 CMe₂): 31 P. δ 27.63.

To a solution of 17 (160 mg, 0.39 mmol) in deuteriochloroform (2.5 mL with 1% of Me₄Si) was added bromotrimethylsilane (150 μ L, 1.1 mmol), and the transesterification was followed by ¹H-n.m.r. spectroscopy. After 1 h, the solution was concentrated, the residue was dissolved in acetonitrile (2 mL) containing 5% of water, which, after 15 min, was concentrated. The residue was dissolved in water, the pH was adjusted to 8 with 0.2M ammonium hydroxide, and the solution was concentrated with methanol to dryness, to give 18 (50 mg, 44%). N.m.r. data (D₂O): ¹H, δ 1.6–2.3 (m, 4 H, H-1,1′,3a,3e). 3.5–4.1 (m, 6 H, H-2,5,6,7,8,8′), 4.3–4.5 (m, 1 H, H-4); ¹³C, δ 32.43 (C-1, ¹ $J_{C,P}$ 131 Hz), 33.32 (C-3, ³ $J_{C,P}$ 10 Hz), 65.14 (C-8), 67.81, 69.27, 71.92, 72.31, 72.92 (C-2.4,5,6,7); ¹³P, δ 21.18 (an additional peak at 21.30). A correct elemental analysis was not obtained. Mass spectrum (f.a.b.): Calc. for C₈H₁₆O₈P (M - NH₄)⁻: m/z 271.0583. Found: m/z 271.0587.

Dimethyl (2,6-anhydro-3-deoxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octonyl) methylphosphonate (19). — Dimethyl methylphosphonate (580 mg, 4.7 mmol) was added to 1.32M butyl-lithium (4.5 mmol) in tetrahydrofuran (30 mL) at -70° . After 8 min, a solution of 4 (640 mg, 2.0 mmol) in tetrahydrofuran (3 mL) was added, and the mixture was allowed to reach room temperature slowly. After neutralisation with ammonium sulphate (2 g) and the addition of water (0.5 mL), followed by stirring for 1 min, the solution was dried (NaHCO₃ and Na₂SO₄), filtered, and concentrated. Column chromatography (acetone-ethyl acetate, 1:1) of the residue gave 19 (536 mg, 64%), $[\alpha]_D = 10^{\circ}$ (c 1.4, dichloromethane), R_F 0.47 (acetone-ethyl acetate, 1:1). N.m.r. data (CDCl₃): 1 H, δ 1.37, 1.42, 1.48 (3 s, 12 H, 2 CMe₂), 1.96 (ddd, 1 H, $J_{3d,3e} = 15.0$, $J_{3a,2}$ 11.4, $J_{3d,4}$ 2.7 Hz, H-3a). 2.18 (ddd, 1 H, $J_{3e,2}$ 5.1, $J_{3e,4}$ 2.7 Hz, H-3e), 3.11–3.61 (ABX, 2 H, CH₂P), 3.69 (dd, 1 H, $J_{6.7}$ 6.8, $J_{6.5}$ 1.7 Hz, H-6), 3.77 (d, 3 H, ${}^{2}J_{Mc,P}$ 3.2 Hz, OMe), 3.83 (d, 3 H, ${}^{2}J_{Mc,P}$ 3.2

Hz, OMe), 4.11–4.30 (m, 3 H, H-7,8,8'), 4.33 (dd, 1 H, $J_{5,4}$ 8.1 Hz, H-5), 4.47 (dd, 1 H, H-2), 4.59 (ddd, 1 H, H-4); ¹³C, δ 24.46, 25.29, 26.13, 26.56, 26.78 (2 CMe₂ and C-3), 36.22 (CH₂, ¹ $J_{C,P}$ 130 Hz), 52.87, 53.15 (2 OMe, ² $J_{C,P}$ 6 Hz), 66.43 (C-8), 69.70, 71.99, 72.21, 74.29, 74.49 (C-2,4,5,6,7), 108.90 (overlapping signals, 2 CMe₂), 204.60 (C-1, ² $J_{C,P}$ 7 Hz); ³¹P, δ 22.58. A correct elemental analysis was not obtained. Mass spectrum (f.a.b.): Calc. for C₁₇H₂₈O₉P (M – H)⁻: m/z 407.1471. Found: m/z 407.1471.

Monocyclohexylammonium (2,6-anhydro-3-deoxy-D-glycero-D-talo-octonyl)-methylphosphonate (20). — Bromotrimethylsilane (0.26 mL, 2.0 mmol) was stirred with a solution of 19 (400 mg, 0.98 mmol) in dichloromethane (3 mL) at room temperature for 20 min. Water (1 mL) was added and, after 1 h, the clear solution was concentrated. The phosphonic acid was converted into the monocyclohexylammonium salt by dissolution in water and addition of cyclohexylamine (110 mg, 1.1 mmol), to give 20 (160 mg, 41%), m.p. 133–134° (from water-ethanol). N.m.r. data (H₂O): 13 C, δ 26.16, 26.68, 32.87, 52.96 (cyclohexyl), 31.79 (C-3), 44.64 (CH₂, 1 J_{C,P} 108 Hz), 65.49 (C-8), 68.76, 71.59, 79.71, 82.19 (C-2,4,5,6,7), 209.85 (C-1, 2 J_{C,P} 6 Hz); 31 P, δ 5.40 (small additional peaks at 5.61, 5.64, and 5.67). A correct elemental analysis was not obtained. Mass spectrum (f.a.b.): Calc. for C₉H₁₆O₉P (M - C₁₆H₁₄N)⁻: m/z 299.0532. Found: m/z 299.0530.

2,6-Anhydro-3-deoxy-4,5:7,8-di-O-isopropylidene-1-O-toluene-p-sulphonyl-D-glycero-D-talo-octitol (21). — Toluene-p-sulphonyl chloride (470 mg, 2.5 mmol) was added to a solution of 14 (264 mg, 0.92 mmol) in dry dichloromethane (20 mL) and pyridine (4.5 mL), which was then heated under reflux overnight. Conventional work-up and column chromatography (ethyl acetate-hexane, 1:1) of the product gave 21 (370 mg, 76%). Recrystallisation from ether-light petroleum gave material with m.p. 82–83°, $[\alpha]_D$ +4° (c 0.5, dichloromethane), R_F 0.50 (ethyl acetate-hexane, 1:1). N.m.r. data (CDCl₃): ${}^{1}H$, δ 1.34, 1.35, 1.38, 1.45 (4 s, 12 H, 2 CMe₂), 1.80–1.86 (m, 2 H, H-3a,3e), 2.45 (s, 3 H, Ts Me), 3.40 (dd, 1 H, $J_{6,7}$ 8.2, $J_{6,5}$ 1.5 Hz, H-6), 3.79–4.20 (m, 6 H, H-1a,1b,2,7,8,8'), 4.33 (dd, 1 H, $J_{5,4}$ 8.2 Hz, H-5), 4.56 (ddd, 1 H, $J_{4,3a}$ 3.0, $J_{4,3e}$ 3.0 Hz, H-4), 7.36 (d, 2 H, aromatic), 7.80 (d, 2 H, aromatic); ${}^{13}C$, δ 21.60 (CH₃), 24.42, 25.30, 26.08, 26.93 (2 CMe₂), 26.61 (C-3), 67.09 (C-8), 72.07 (C-1), 66.21, 69.76, 72.07, 72.24, 73.97 (C-2,4,5,6,7), 108.85, 109.22 (2 CMe₂), 127.92, 129.92, 132.96, 144.93 (aromatic).

Anal. Calc. for $C_{21}H_{30}O_8S$: C, 57.0; H, 6.8; S, 7.2. Found: C, 56.9; H, 6.9; S, 7.1.

3,7-Anhydro-2,4-dideoxy-5,6:8,9-di-O-isopropylidene-D-glycero-D-talo-nononitrile (22). — A mixture of 21 (270 mg, 0.61 mmol), potassium cyanide (200 mg, 3.1 mmol), and N,N-dimethylformamide (20 mL) was heated at 100° for 1.5 h and then concentrated. Column chromatography (ethyl acetate-hexane, 1:2) of the residue gave 22 (160 mg, 88%), m.p. 89-90°, $[\alpha]_D$ +3° (c 1.7, dichloromethane), R_F 0.67 (ethyl acetate). N.m.r. data (CDCl₃): 1 H, δ 1.37, 1.38, 1.41, 1.50 (4 s, 12 H, 2 CMe₂), 1.8-2.1 (m, 2 H, H-4a,4e), 2.4-2.7 (ABX, 2 H, CH₂), 3.56 (dd, 1 H, $J_{7.8}$ 8.1, $J_{7.6}$ 1.5 Hz, H-7), 4.0-4.3 (m, 4 H, H-6,8,9,9'), 4.40 (dd, 1 H, $J_{6.5}$ 8.2 Hz,

H-6), 4.60 (ddd, 1 H, $J_{5,4a}$ 2.7, $J_{5,4e}$ 2.7 Hz, H-5); ¹³C, δ 24.28, 25.30, 26.08, 26.88 (2 CMe_2), 24.81 (C-2), 30.09 (C-4), 64.29 (C-3), 67.09 (C-9), 69.67, 71.81, 72.10, 73.97 (C-5,6,7,8), 108.90, 109.29 (2 CMe_2), 117.35 (C-1).

Anal. Calc. for $C_{15}H_{23}NO_5$; C, 60.6; H, 7.8; N, 4.7. Found: C, 60.35; H, 7.8; N, 4.5.

Methyl 3,7-anhydro-2,4-dideoxy-D-glycero-D-talo-nononate (23). — To a solution of 22 (520 mg, 1.75 mmol) in anhydrous methanol (15 mL) was added chlorotrimethylsilane (0.91 mL, 7.2 mmol) at -10°, and dry HCl was bubbled through the solution for 10 min. The mixture was then stirred for 2.5 h at -20° , the temperature was raised slowly to 0° , and water (1 mL) was added dropwise. The pH was adjusted to 5.5 with M sodium hydroxide and the solution was then concentrated with methanol to dryness. A solution of the residue in methanol was neutralised with Amberlite IR-45 (CO₃²) resin and then passed over Dowex 50W-X8 (H⁻) resin with methanol as eluent. Concentration of the eluate gave 23 (270 mg, 63%), which, after recrystallisation from methanol-ether, had m.p. 113-117°, $[\alpha]_D$ +124° (c 0.7, methanol), R_E 0.25 (ethyl acetate-methanol-toluene, 7:2:1); $\nu_{\text{max}}^{\text{KBr}}$ 1730 cm⁻¹ (ester). N.m.r. data (CD₃OD): ¹H, δ 1.57 (dd, 1 H, $J_{4c,4a}$ –13, $J_{4c,5}$ 4.5 Hz, H-4e), 2.09 (ddd, 1 H, $J_{4a.5}$ 13, $J_{4a.3}$ 6 Hz, H-4a), 2.45-2.93 (ABX, 2 H, H-2a,2b), 3.42–3.95 (m, 9 H, H-5,6,7,8,9,9', OMe), 4.45 (ddd, 1 H, $J_{3,2b}$ 10, $J_{3,2a}$ 5 Hz, H-3); 13 C, δ 32.13 (C-4), 37.61 (C-2), 52.25 (OMe), 65.21 (C-9), 67.09, 68.64. 71.27, 73.24 (C-3,5,6,7,8), 173.56 (C-1).

Anal. Calc. for $C_{10}H_{18}O_7 \cdot 0.25 H_2O$: C, 47.15; H, 7.3. Found: C, 47.0; H, 7.0. Ammonium 3,7-anhydro-2,4-dideoxy-D-glycero-D-talo-nononate (24). — A solution of 23 (173 mg, 0.69 mmol) in M sodium hydroxide (3 mL) was stored for 30 min, and then eluted from Dowex 50W-X8 (NH $_4^+$) resin with water. Concentration of the cluate gave 24 (125 mg, 70%), $[\alpha]_D$ +67° (c 1, water), R_F 0.22 (chloroform-methanol-water, 10:10:1). N.m.r. data (D₂O): 1H , δ 1.64 (dd, 1 H, $J_{4e,4a}$ -13.5, $J_{4e,5}$ 2.5 Hz, H-4e), 2.02 (ddd, 1 H, $J_{4a,5}$ 13, $J_{4a,3}$ 6 Hz, H-4a), 2.33–2.91 (ABX, 2 H, H-2a,2b), 3.45–4.11 (m, 6 H, H-5,6,7.8,9,9'), 4.47 (ddd, 1 H, $J_{3,2b}$ 10, $J_{3,2a}$ 5 Hz, H-3); ^{13}C , δ 32.42 (C-4), 40.13 (C-2), 62.10 (C-9), 64.14, 65.14, 67.56, 68.86, 69.08 (C-3,5,6,7,8), 164.77 (C-1). Mass spectrum (f.a.b.): Calc. for $C_9H_{15}O_7$ (M - NH $_4$) $^{-1}$: m/z 235.0818. Found: m/z 235.0811.

Methyl N-acetyl-2,6-anhydro-3-deoxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octonamide (26). — Compound 4 (500 mg, 1.58 mmol) was stirred with 2M NaOH (5 mL) for 2 h at room temperature, and the solution was passed through Dowex 50W-X8 (Et₃NH) resin and concentrated to afford 25 (497 mg, 78%), m.p. $100-103^{\circ}$, [α]_D -42° (c 1, chloroform), R_F 0.38 (ether). N.m.r. data (CDCl₃): 1 H, δ 1.26 (t, Me), 1.36, 1.37, 1.40, 1.49 (4 s, 12 H, 2 CMe₂), 1.89 (ddd, 1 H, $J_{3a,3e}$ = 15, $J_{3a,2}$ 11, $J_{3a,4}$ 3 Hz, H-3a), 2.33 (ddd, 1 H, $J_{3e,2}$ 6, $J_{3e,4}$ 3.2 Hz, H-3e), 3.09 (q, CH₂), 3.56 (dd, 1 H, $J_{6,7}$ 8, $J_{6,5}$ 1.5 Hz, H-6), 4.06-4.36 (m, 4 H, H-5,7,8,8′), 4.44 (dd, 1 H, H-2), 4.58 (ddd, 1 H, $J_{4,5}$ 8 Hz, H-4), 9.17 (bs, NH); 13 C, δ 8.46 (CH₃), 25.14, 26.37, 27.08 (2 CMe₂), 27.52 (C-3), 44.91 (CH₂), 67.30 (C-8), 69.58 (C-2), 70.42 (C-4), 72.39, 72.45 (C-5,6), 74.06 (C-7), 108.99, 109.14 (2 CMe₂), 177.71 (C-1). This compound was used in the next step without further characterisation.

Triethylamine (0.15 mL, 1.1 mmol) and ethyl chloroformate (50 μ L, 0.54 mmol) were added to a suspension of 25 (180 mg, 0.45 mmol) in N, N-dimethylformamide (5 mL) at -10°. After 5 min, a solution of glycine methyl ester hydrochloride (85 mg, 0.68 mmol) and triethylamine (0.15 mL) in N, N-dimethylformamide (2 mL) was added dropwise, and stirring was continued for 1 h. The mixture was then filtered and concentrated. Column chromatography (ethyl acetatechloroform, 1:1) of the residue afforded a pale-yellow syrup which was filtered through silica gel with ether to give **26**, isolated as a colourless glass (140 mg, 82%), $[\alpha]_D$ -26° (c 1.6, chloroform), R_F 0.58 (ethyl acetate-chloroform, 1:1). N.m.r. data (CDCl₃): 1 H, δ 1.28, 1.32, 1.37, 1.40 (4 s, 12 H, 2 CMe₂), 1.71 (ddd, 1 H, $J_{3a,3e}$ -15.5, $J_{3a,2}$ 11.8, $J_{3a,4}$ 2.5 Hz, H-3a), 2.42 (ddd, 1 H, $J_{3e,2}$ 5.5, $J_{3e,4}$ 2.8 Hz, H-3e), 3.63 (dd, 1 H, $J_{6.7}$ 4.8, $J_{6.5}$ 1.5 Hz, H-6), 3.69 (s, 3 H, OMe), 3.95–4.26 (m, 6 H, H-5,7,8,8', CH_2), 4.3-4.65 (dd, 1 H, H-2; ddd, 1 H, H-4), 7.30 (t, 1 H, NH); ^{13}C , δ 24.53, 25.42, 25.97, 26.52 (2 CMe₂), 27.22 (C-3), 40.73 (CH₂), 52.29 (OMe), 65.70 (C-8), 69.34 (C-2), 70.14 (C-4), 71.64 (C-6), 72.23 (C-7), 75.18 (C-5), 108.57, $109.17 (2 \text{ CMe}_2), 169.94, 173.23 (C-1, methyl ester C=0).$

Anal. Calc. for $C_{17}H_{27}NO_8 \cdot 0.75 H_2O$: C, 52.8; H, 7.4; N, 3.6. Found: C, 52.8; H, 7.0; N, 3.4.

Ammonium N-acetyl-2,6-anhydro-3-deoxy-D-glycero-D-talo-octonamide (27). — Compound 26 (100 mg, 0.27 mmol) was deprotected according to the standard procedure. After concentration, the residue was triturated with ether and pentane, to afford the ester as a white semi-crystalline residue (60 mg, 77%), $R_{\rm F}$ 0.52 (ethyl acetate-methanol-water, 7:2:1). N.m.r. data (CD₃OD): 1 H, δ 2.04 (ddd, 1 H, $J_{3a,3e}$ –12, $J_{3a,4}$ 12, $J_{3a,2}$ 6 Hz, H-3a), 2.32 (ddd, 1 H, $J_{3e,4}$ 5, $J_{3e,2}$ 1.5 Hz, H-3e), 2.92 (d, 2 H, CH₂), 3.50–4.05 (m, 9 H, H-4,5,6,7,8,8', including OMe at δ 3.72), 4.48 (dd, 1 H, H-2); 13 C, δ 28.23 (C-3), 41.69 (CH₂), 52.69 (OCH₃), 64.86 (C-8), 67.94 (overlapping signals, C-4, C-5), 70.42 (C-7), 74.86 (C-6), 75.85 (C-2), 171.65, 174.37 (amide, ester).

The ester (60 mg, 0.20 mmol) was treated with 0.5M NaOH (5 mL) for 1 h and the residue was passed over Dowex 50W-X8 (NH₄⁺) resin, and then concentrated to give **27** (60 mg, 100%) as a semi-crystalline residue, $[\alpha]_D$ +51.5° (c 1.2, methanol), R_F 0.19 (chloroform-methanol-water, 10:10:1). N.m.r. data (D₂O): ¹H, δ 2.00 (ddd, 1 H, $J_{3a,3e}$ -12, $J_{3a,4}$ 12, $J_{3a,2}$ 6 Hz, H-3a), 2.30 (ddd, 1 H, $J_{3e,4}$ 5, $J_{3e,2}$ ~1 Hz, H-3e), 3.58-3.96 (m, 8 H, H-4,5,6,7,8,8', CH₂), 4.61 (dd, 1 H, H-2); ¹³C, δ 28.91 (C-3), 45.61 (CH₂), 65.45 (C-8), 68.69, 68.54 (C-4,5), 71.43 (C-7), 75.92, 76.57 (C-2,6), 175.22, 178.96 (C-1, carboxylate). Mass spectrum (f.a.b.): Calc. for C₁₀H₁₆NO₈ (M - NH₄)⁻: m/z 278.0876. Found: m/z 278.0869.

2,6-Anhydro-N-benzyl-3-deoxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octonamide (28). — The amidation of 25 (280 mg, 0.69 mmol) in acetonitrile (5 mL) with triethylamine (140 μ L), ethyl chloroformate (67 μ L, 0.7 mmol), and benzylamine (110 μ L, 1 mmol) was performed as described for 26. Column chromatography (ether) of the product gave 28 (250 mg, 92%), m.p. 93–94°, [α]_D -61° (c 1.1, chloroform), R_F 0.60 (ether). N.m.r. data (CDCl₃): 1 H, δ 1.12, 1.28,

1.34, 1.45 (4 s, 12 H, 2 CMe₂), 1.78 (ddd, 1 H, $J_{3a,3e}$ –15.5, $J_{3a,2}$ 11.6, $J_{3a,4}$ 2.6 Hz, H-3a), 2.57 (ddd, 1 H, $J_{3e,2}$ 6, $J_{3e,4}$ 3 Hz, H-3e), 3.70 (dd, 1 H, $J_{6,7}$ 3.5, $J_{6,5}$ 1.5 Hz, H-6), 3.94–4.30 (m, 4 H, H-5,7,8,8'), 4.46–4.70 (m, 4 H, H-2,4, CH₂), 7.3 (s, 5 H, Ph); ¹³C, δ 24.53, 25.32, 25.92 (2 CMe₂), 26.92 (C-3), 43.17 (CH₂), 64.96 (C-8), 69.44 (C-2), 70.39 (C-4), 71.24 (C-6), 72.33 (C-7), 75.47 (C-5), 108.03, 109.27 (2 CMe₂), 127.52, 127.97, 128.62, 137.79 (aromatic), 172.64 (C-1).

Anal. Calc. for $C_{21}H_{29}NO_6 \cdot 0.25 H_2O$: C, 63.7; H, 7.5; N, 3.5. Found: C, 63.5; H, 7.4; N, 3.5.

2,6-Anhydro-N-benzyl-3-deoxy-D-glycero-D-talo-octonamide (29). — Deprotection of 28 (200 mg, 0.51 mmol) by the general procedure and crystallisation of the product from methanol-ether gave 29 (110 mg, 69%), m.p. 155–156°. $[\alpha]_D$ +49° (c 0.4, methanol), R_F 0.60 (ethyl acetate-methanol-toluene, 7:2:1). N.m.r. data (CD₃OD): 1 H, δ 2.04 (ddd, 1 H, $J_{3a,3v}$ –12, $J_{3a,4}$ 12, $J_{3a,2}$ 6 Hz. H-3a), 2.35 (ddd, $J_{3e,4}$ 6, $J_{3e,2}$ 2 Hz, H-3e), 3.44–3.90 (m, 6 H, H-4,5,6,7,8.8'), 4.33–4.57 (m. 3 H, H-2, CH₂), 7.28 (s. 5 H, Ph); 13 C, δ 28.22 (C-3), 44.02 (CH₂), 64.71 (C-8), 67.85, 68.10 (C-4,5), 70.29 (C-7), 75.08 (C-6), 75.92 (C-2), 128.22, 128.67, 129.51, 139.38 (aromatic), 178.28 (C-1).

Anal. Calc. for $C_{15}H_{21}NO_6$: C, 57.9; H, 6.8; N, 4.5. Found: C, 58.1; H, 6.7; N, 4.3.

2,6-Anhydro-3-deoxy-N-hexyl-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octonamide (30). — Compound 30 was prepared from 25 (250 mg, 0.62 mmol) in tetrahydrofuran (5 mL) by reaction with triethylamine (0.2 mL), ethyl chloroformate (66 μ L, 0.68 mmol), and hexylamine (0.12 mL, 0.62 mmol) as described for 26. Column chromatography (ethyl acetate-chloroform, 1:1) of the product gave 30 (170 mg, 71%). which crystallised after prolonged storage and had m.p. 33–35°, [α]_D -38° (c 1.9, chloroform), R_F 0.65 (ethyl acetate-chloroform, 1:1). N.m.r. data (CDCl₃): 1 H, δ 0.82 (t, 3 H, H-6',6',6'), 1.19–1.39 (m. 20 H, 2 CMe₂ and H-2',2',3',3',4',4',5',5'), 1.65 (ddd, 1 H, $J_{3u,3e}$ -15.5, $J_{3u,2}$ 11.7. $J_{3u,4}$ 2.5 Hz, H-3a), 2.45 (ddd, 1 H, $J_{3e,2}$ 6, $J_{3e,4}$ 3 Hz, H-3e), 3.19 (q, 2 H, H-1.1), 3.61 (dd, 1 H, $J_{6,7}$ 4, $J_{6,5}$ 1.5 Hz, H-6), 3.94–4.20 (m, 4 H, H-5,7.8,8), 4.39 (dd, 1 H, H-2), 4.52 (ddd, 1 H, $J_{4,5}$ 8 Hz, H-4), 6.9 (t, 1 H, NH); 13 C, δ 14.01 (C-6'), 22.53 (C-5'), 24.53, 25.42, 25.92, 26.47 (2 CMe₂), 26.57, 27.02, 29.56 (C-3,3',4'), 31.46 (C-2'), 38.93 (C-1'), 65.16 (C-8), 69.34 (C-2), 70.34 (C-4), 71.24 (C-6), 72.38 (C-7), 75.52 (C-5), 108.13, 109.22 (2 CMe₂), 172.58 (C-1).

Anal. Calc. for $C_{20}H_{35}NO_6$: C, 62.3; H, 9.15; N, 3.6. Found: C, 62.3; H, 9.3; N, 3.4.

2,6-Anhydro-3-deoxy-N-hexyl-D-glycero-D-talo-octonamide (31). — Compound 30 (170 mg, 0.44 mmol) was deprotected according to the standard procedure. Crystallisation of the product from methanol—ether afforded 31 (120 mg, 92%), m.p. 128–129°, $[\alpha]_D$ +59° (c 1, methanol), R_F 0.68 (ethyl acetate—methanol—toluene, 7:2:1). N.m.r. data (CD₃OD): 1 H, δ 0.90 (t, 3 H, H-6′,6′,6′), 1.23–1.70 (m, 8 H, H-2′,2′,3′,3′,4′,4′,5′,5′), 2.00 (ddd, 1 H, $J_{3a,3e}$ –12, $J_{3a,4}$ 12, $J_{3a,2}$ 6 Hz, H-3a), 2.30 (ddd, 1 H, $J_{3e,4}$ 5, $J_{3e,2}$ 2 Hz, H-3e), 3.10–4.00 (m, 8 H, H-

1',1',4,5,6,7,8,8), 4.39 (dd, 1 H, H-2); ¹³C, δ 14.41 (C-6'), 23.68 (C-5'), 27.82, 28.12, 30.31 (C-3,3',4'), 32.65 (C-2'), 40.43 (C-1'), 64.66 (C-8), 67.10, 67.85 (C-4,5), 70.24 (C-7), 75.03 (C-6), 75.72 (C-2), 173.03 (C-1).

Anal. Calc. for $C_{14}H_{27}NO_6 \cdot 0.25 H_2O$: C, 54.3; H, 8.9; N, 4.5. Found: C, 54.2; H, 8.8; N, 4.6.

Benzyl 2,6-anhydro-3-deoxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octonate (32). — Compound 4 (600 mg, 1.9 mmol) was heated at 120° with benzyl alcohol (20 mL) and a catalytic amount of magnesium methoxide for 7 h under nitrogen. The solution was neutralised with Amberlite IR-120 (H+) resin (prewashed with methanol and dried), filtered, and concentrated. Column chromatography (ethyl acetate-toluene, 1:9) of the product gave 32 (600 mg, 81%), isolated as a colourless syrup, $[\alpha]_D$ -26° (c 1.2, dichloromethane), R_F 0.57 (ethyl acetate-toluene, 1:2). N.m.r. data (CDCl₃): 1 H, δ 1.30, 1.37, 1.40, 1.48 (4 s, 12 H, 2 CMe₂), 1.85 (ddd, 1 H, $J_{3a,3e}$ -15, $J_{3a,2}$ 11.7, $J_{3a,4}$ 2.4 Hz, H-3a), 2.31 (ddd, 1 H, $J_{3e,2}$ 5.9, $J_{3e,4}$ 3.4 Hz, H-3e), 3.51 (dd, 1 H, $J_{6,7}$ 8.3, $J_{6,5}$ 1.5 Hz, H-6), 4.03-4.26 (m, 3 H, H-7,8,8'), 4.33 (dd, $J_{5,4}$ 7.8 Hz, H-5), 4.55-4.63 (m, 2 H, H-2,4), 5.18 (s, 2 H, CH₂), 7.35 (s, 5 H, Ph); 13 C, δ 24.96, 25.20, 26.22, 26.85 (2 CMe₂), 27.00 (C-3), 66.60 (CH₂), 67.23 (C-8), 68.52 (C-2), 69.76 (C-4), 72.32 (C-5), 72.90 (C-6), 73.78 (C-7), 109.29 (overlapping signals, 2 CMe₂), 128.24, 128.29, 128.61, 135.61 (aromatic), 172.58 (C-1).

Anal. Calc. for C₂₁H₂₈O₇: C, 64.3; H, 7.2. Found: C, 64.6; H, 6.8.

Benzyl 2,6-anhydro-3-deoxy-D-glycero-D-talo-octonate (33). — Compound 32 (580 mg, 1.48 mmol) was deprotected according to the standard procedure. Column chromatography (ethyl acetate-methanol-toluene, 7:2:1) of the product gave 33 (210 mg, 45%). Recrystallisation from methanol gave material with m.p. 131–132°, $[\alpha]_D$ +60° (c 1.15, methanol), R_F 0.35 (ethyl acetate-methanol-toluene, 7:2:1). N.m.r. data (CD₃OD): 1 H, δ 2.12–2.19 (m, 2 H, H-3a,3e), 3.46–3.60 (m, 3 H), 3.73–3.80 (m, 2 H), 3.92 (d, 1 H), 4.60 (dd, 1 H), 5.20 (AB, 2 H, CH₂), 7.38 (s, 5 H, Ph); 13 C, δ 29.70 (C-3), 65.68 (C-8), 68.01, 68.11, 71.25, 73.95, 76.53 (C-2,4,5,6,7), 129.51, 129.70, 137.24 (aromatic), 173.29 (C-1).

Anal. Calc. for $C_{15}H_{20}O_7 \cdot 0.5$ CH_3OH : C, 56.7; H, 6.75. Found: C, 56.8; H, 6.4.

Hexyl 2,6-anhydro-3-deoxy-4,5:7,8-di-O-isopropylidene-D-glycero-D-talo-octonate (34). — Transesterification of 4 (430 mg, 1.36 mmol) in 1-hexanol (17 mL) with magnesium methoxide was carried out as for 32. Column chromatography (ethyl acetate-toluene, 1:6) of the product gave pure 34 (470 mg, 90%) as a colourless syrup, $[\alpha]_D$ –33° (c 1.55, chloroform), R_F 0.58 (ethyl acetate-toluene, 1:2). N.m.r. data (CDCl₃): 1 H, δ 0.89 (t, 3 H, H-6',6',6'), 1.2–1.7 (m, 20 H, 2 CMe₂ and H-2',2',3',3',4',4',5',5'), 1.85 (ddd, 1 H, $J_{3a,3e}$ –15, $J_{3a,2}$ 11.7, $J_{3a,4}$ 2.4 Hz, H-3a), 2.31 (ddd, 1 H, $J_{3e,2}$ 5.9, $J_{3e,4}$ 3.2 Hz, H-3e), 3.51 (d, 1 H, $J_{6,7}$ 8.0 Hz, H-6), 4.10–4.25 (m, 5 H, H-1',1',7,8,8), 4.35 (dd, 1 H, $J_{5,6}$ 1.5, $J_{5,4}$ 7.8 Hz, H-5), 4.5–4.6 (m, 2 H, H-2,4): 13 C, δ 13.99 (C-6'), 22.55 (C-5'), 25.61, 26.30, 28.68 (C-3,3',4'), 25.03, 25.25, 26.98, 27.10 (2 CMe₂), 31.45 (C-2'), 65.14 (C-1'), 67.33 (C-8), 68.57 (C-2), 69.91 (C-4), 72.42 (C-5), 72.95 (C-6), 73.88 (C-7), 109.34, 109.41 (2 CMe₂), 172.98 (C-1).

Anal. Calc. for C₂₀H₃₄O₇: C, 62.15; H, 8.9. Found: C. 61.9; H. 9.0.

Hexyl 2,6-anhydro-3-deoxy-D-glycero-D-talo-octonate (35). — Compound 34 was deprotected according to the standard procedure. Column chromatography (ethyl acetate-toluene-methanol, 7:1:1) of the product gave 35 (270 mg, 73%). Recrystallisation from ethanol-ether gave material with m.p. 114-116°, $[\alpha]_D$ +59° (c 0.1, methanol), R_{+} 0.36 (ethyl acetate-methanol-toluene, 7.2:1). N.m.r. data (CD₃OD): 1 H, δ0.92 (t, 3 H, H-6',6',6'), 1.25-1.8 (m, 8 H, H-2',2',3',3',4',4',5',5'), 2.1-2.2 (m, 2 H, H-3a,3e), 3.50 (d, 1 H, H-6), 3.48-3.83 (m, 4 H, H-4,7.8,8), 3.96 (d, 1 H, H-5), 4.16 (m, 2 H, OCH₂), 4.55 (dd, 1 H, H-2): 13 C, δ 14.33 (C-6'), 23.55 (C-5'), 26.68, 29.65 (C-3.3',4'), 32.52 (C-2'), 65.65 (C-1'), 66.48 (C-8), 68.04, 71.08, 73.83, 76.38 (C-2,4,5,6,7), 173.51 (C-1).

Anal. Calc. for C₁₄H₂₆O₇; C, 54.9; H, 8.6. Found: C. 54.8; H. 8.3.

ACKNOWLEDGMENTS

We thank Miss K. Crona and Miss S. Wingborg for technical assistance, Dr. K. Gustafsson for the enzyme inhibition tests, Dr. L. Magni for the MIC tests, Professor Lennart Kenne for running the 400-MHz spectrum, and the National Swedish Board for Technical Development for financial support

REFERENCES

- 1 E. C. Heath and M. A. Ghalambor, Biochem Biophys. Res. Commun., 10 (1963) 340-345.
- 2 F. M. UNGER, Adv. Carbohydr. Chem. Biochem., 38 (1981) 323-388
- 3 M. J. OSBORN, Proc. Natl. Acad. Sci. U S.A., 50 (1963) 499-506.
- 4 E. C. HEATH, R. M. MAYER, R. D. EDSTROM, AND C. A. BEAUDREW. Ann. N.Y. Acad. Sci., 133 (1966) 315-333.
- 5 D. C. Ellwood, J. Gen. Microbiol., 60 (1970) 373-380.
- 6 S. S. GOLDBERG, M. N. CORDEIRO, A. A. SILVA PERFIRA, AND M. L. MARES-GUIA, Int. J. Parasitol., 13 (1983) 11–18.
- W. S. YORK, A. G. DARVILL, M. MCNEIL, and P. Albersheim, Carbohydr. Res., 438 (1985) 109-126.
- 8 P. W. TAYLOR, Biochem. Biophys. Res. Commun., 61 (1974) 148-154
- 9 A. K. BHATTACHARJEE, H. J. JENNINGS, AND C. P. KENNY, Biochem. Biophys. Res. Commun., 61 (1974) 489-493.
- 10 W. F. VANN AND K. JANN, Infect. Immun., 25 (1979) 85-92.
- 11 P. MESSNER AND F. M. UNGER, Biochem. Biophys. Res. Commun., 96 (1980) 1003-1010.
- 12 H. J. Jennings, K.-G. Rosell, and K. G. Johnson, Carbohydr. Res., 105 (1982) 45-56.
- 13 B. JANN, P. HOFMANN, AND K. JANN, Carbohydr. Res., 120 (1983) 131-141
- 14 M. A. SCHMIDT AND K. JANN, Eur. J. Biochem., 131 (1983) 509-517.
- 15 K. Jann, ACS Symp. Ser., 231 (1983) 171-191.
- 16 P. D. RICK AND M. J. OSBORN, Proc. Natl. Acad. Sci. U.S.A., 69 (1972) 3756-3760.
- 17 P. D. RICK AND M. J. OSBORN, J. Biol. Chem., 252 (1977) 4895–4903
- 18 M. A. GHALAMBOR AND E. C. HEATH, J. Biol. Chem., 241 (1966) 3216-3221.
- 19 P. H. RAY, C. D. BENEDICT, AND H. GRASMUK, J. Bacteriol., 145 (1981) 1273-1280.
- 20 M. A. GHALAMBOR AND E. C. HLATH, Biochem. Biophys. Res. Commun., 10 (1963) 346-351.
- 21 P. H. RAY, J. E. KELSFY, E. C. BIGHAM, C. D. BENFDICT, AND T. A. MILLER, ACS Symp. Sci., 231 (1983) 141–169.
- 22 A. CLAESSON, K. LUTHMAN, K. GUSTAFSSON, AND G. BONDESSON, Biochem. Biophys. Res. Commun., 143 (1987) 1063-1068.

- 23 W. E. KOHLBRENNER AND S. W. FESIK, J. Biol. Chem., 260 (1985) 14695-14700.
- 24 S. M. STRAIN, S. W. FESIK, AND I. M. ARMITAGE, J. Biol. Chem., 258 (1983) 2906-2910.
- 25 S. M. STRAIN, S. W. FESIK, AND I. M. ARMITAGE, J. Biol. Chem., 258 (1983) 13466-13477.
- 26 H. BRADE, U. ZÄHRINGER, E. T. RIETSCHEL, R. CHRISTIAN, G. SCHULZ, AND F. M. UNGER, Carbohydr. Res., 134 (1984) 157-166.
- 27 K. LUTHMAN, M. ORBE, T. WAGLUND, AND A. CLAESSON, J. Org. Chem., in press.
- 28 A. K. BHATTACHARJEE, H. J. JENNINGS, AND C. P. KENNY, Biochemistry, 17 (1978) 645-651.
- 29 S. REIFFERS, H. WYNBERG, AND J. STRAITING, Tetrahedron Lett., (1971) 3001-3004.
- 30 C. E. McKenna and J. Schmidhauser, J. Chem. Soc., Chem. Commun., (1979) 739.
- 31 W. G. DAUBEN, G. H. BEASLEY, M. D. BROADHURST, B. MULLER, D. J. PEPPARD, P. PESNELLE, AND C. SUTER, J. Am. Chem. Soc., 97 (1975) 4973-4980.
- 32 B. BIRDSALL, N. J. M. BIRDSALL, AND J. FEENEY, J. Chem. Soc., Chem. Commun., (1972) 316-317.
- 33 M. L. MARTIN, G. J. MARTIN, AND J.-J. DELPUECH, Practical NMR Spectroscopy, Heyden, London, 1980, pp. 212-213.
- 34 J. HAVERKAMP, T. SPOORMAKER, L. DORLAND, J. F. G. VLIEGENTHART, AND R. SCHAUER, J. Am. Chem. Soc., 101 (1979) 4851–4853.
- 35 F. M. UNGER, D. STIX, AND G. SCHULZ, Carbohydr. Res., 80 (1980) 191-195.
- 36 C. CONE AND L. HOUGH, Carbohydr. Res., 1 (1965) 1-9.
- 37 H. LIBERT, I. SCHUSTER, AND L. SCHMID, Chem. Ber., 101 (1963) 1902-1909.
- 38 J. W. Krajewski, P. Gluzinski, Z. Urbanczyk-Lipkowska, A. Zamojski, and P. Luger, Carbohydr. Res., 139 (1985) 55-63.
- 39 F. H. CANO, C. FOCES-FOCES, M. BERNABE, J. JIMENEZ-BARBERO, M. MARTIN-LOMAS, AND S. PENADES-ULLATE, *Tetrahedron*, 41 (1985) 3875–3886.
- 40 Y. D. KARKHANIS, J. Y. ZELTNER, J. J. JACKSON, AND D. J. CARLO, Anal. Biochem., 85 (1978) 595-601.
- 41 G. BONDESSON, I. FAGERVALL, T. GOSZTONYI, A. JANSSON, AND A. OHLBERGER, J. Labelled Compd. Radiopharmaceuticals, 19 (1982) 239–246.
- 42 A. L. BARRY, The Antimicrobic Susceptibility Test: Principles and Practice, Lea Febiger, Philadelphia, 1976, Ch. 6.
- 43 W. C. STILL, M. KAHN, AND A. MITRA, J. Org. Chem., 43 (1978) 2923-2925.
- 44 W. G. KOFRON AND L. M. BACLAWSKI, J. Org. Chem., 41 (1976) 1879-1880.